

Supplementary Material

1 Supplementary Tables

Table S1. Sampling scheme showing the samples used for each type of analysis.

Analysis Date	HPLC-MS	qPCR	16S rRNA gene amplicon seq.	<i>intI1</i> gene cassette
7 th Dec 20	✓	\checkmark	\checkmark	✓
4 th Jan 21	✓	✓		✓
11 th Jan 21	✓	✓	\checkmark	✓
18 th Jan 21	✓	✓		✓
25 th Jan 21	✓	✓		✓
1 st Feb 21	✓	\checkmark	\checkmark	\checkmark
24 th Feb 21		\checkmark		
25 th Feb 21		\checkmark		
26 th Feb 21		\checkmark	\checkmark	\checkmark
27 th Feb 21		\checkmark		
28 th Feb 21		\checkmark		\checkmark
1 st Mar 21		\checkmark		
2 nd Mar 21		\checkmark		
3 rd Mar 21		\checkmark		
4 th Mar 21	\checkmark	\checkmark	\checkmark	\checkmark

Table S2. Multiple reaction monitoring (MRM) conditions for the analysis of SMX, SDZ and SMZ.

Analyte	Q1 (m/z)	Q3 (m/z)	Declustering Potential DP (V)	Entrance Potential EP (V)	Collision Energy CE (V)	Cell Exit Potential CXP (V)
SMX-d4	258.2	160.0	55	10	22	12
SDZ	251.1	156.1	45	10	22	14
		92.0			37	10
SMZ	279.0	185.9	60	10	25	8
		124.1			32	8
SMX	254.0	156.0	55	10	22	12
	254.0	108.0	55	10	32	10

Gene	Primers*	Sequence (5'-3')	NCBI Ref.Seq.	Amplicon size (bp)	Reference	
	27F	AGAGTTTGATCMTGGCTCAG		1465	Lane D. J., 1991	
	1492R	CGGTTACCTTGTTACGACTT	-		Turner et al., 1999	
	519qF	CCAGCAGCCGCGGTAATAC		410		
	909qR	CCGTCAATTCCTTTRAGTTT		410		
	341F-TS	ACACTCTTTCCCTACACGACGCT CTTCCGATCT CCTACGGGNGGC	- 194			
		WGCAG				
	518R-TS	GACTGGAGTTCAGACCTGTGCTC TTCCGATCT WTTACCGCGGCTG CTGG				
	sul1qF	TGTCGAACCTTCAAAAGCTG	WP_00025			
sul1	sullqR	TGGACCCAGATCCTTTACAG	9031.1	113	Wang et al., 2014	
au 12	sul2qF	ATCTGCCAAACTCGTCGTTA	WP_00104	89	Wang et al., 2014	
SUI2	sul2 sul2qR	CAATGTGATCCATGATGTCG	3260.1	07		
	5'CS	AAACGGATGAAGGCACGAAC	WP_00084	voriable	Frank et al., 2007	
intI1	3°CS	ATTGCGATAACAAGAAAAAGCC	5048.1	variable	171alik et al., 2007	
	intI1qF	CGAACGAGTGGCGGAGGGTG		312	Gillings et al., 2015	
	intI1qR	TACCCGAGAGCTTGGCACCCA				

Table S3. Overview of primers used in this study.

*q in primer name indicates the use in qPCR

Table S4. Site-specific phylotypes and phylotypes introduced by wastewater discharge on a family level (bold face). Only phylotypes with a frequency over 1 % were considered for the analysis.

Site 1	Site 2	Site 3	Site 4	Site 5	Site 6
Caulobacteraceae	Flavobacteriaceae	Rhodobacteraceae			
Nocardiaceae		Diplorickettsiaceae			
Uncl. Rhizobiales		Saprospiraceae			
		Peptostreptococcaceae			

Site 3	Site 4	Site 5	Site 6
aadA1 ^{*1}	aadA1 ^{*1}	$aadA2^{*1}$	$aadA2^{*1}$
$aadA1^{*1}$	$aadA1^{*1}$	aadA6/aadA10 ^{*1}	$aadA2^{*1}$
$aadA1^{*1}$	$aadA1^{*1}$		$aadA2^{*1}$
$aadA11^{*1}$	$aadA11^{*1}$		$bla_{\text{OXA-36}}^{*3}$
$aadA11^{*1}$	$aadA11^{*1}$		
$aadA2^{*1}$	aadA11 ^{*1}		
$aadA2^{*1}$	$aadA2^{*1}$		
$aadA23^{*1}$	$aadA2^{*1}$		
$aadA24^{*1}$	$aadA2^{*1}$		
$aadA25^{*1}$	$aadA2^{*1}$		
$aadA4^{*1}$	$aadA2^{*1}$		
aadA6/aadA10 ^{*1}	$aadA2^{*1}$		
aadA6/aadA10 ^{*1}	$aadA23^{*1}$		
aadA6/aadA10 ^{*1}	$aadA24^{*1}$		
$bla_{\text{BEL-1}}^{*2}$	aadA6/aadA10 ^{*1}		
$bla_{\text{BEL-1}}^{*2}$	$bla_{\text{BEL-1}}^{*2}$		
$bla_{\text{GES-11}}^{*4}$	$bla_{\text{OXA-33}}^{*3}$		
bla_{OXA-33}^{*3}	$bla_{\text{OXA-36}}^{*3}$		
$bla_{\text{OXA-36}}^{*3}$	ereA2 ^{*7}		
$bla_{OXA-392}^{*3}$	qacL		
bla _{OXA-4}	-		
$bla_{\text{OXA-824}}$ *3			
cmlA5 ^{*5}			
$dfrA7^{*6}$			
ereA2 ^{*7}			
$qacL^{*8}$			

Table S5. List of gene cassette inserts found for each site.

Gene family | Drug class

*1 ANT(3") aminoglycoside nucleotidyltransferase | aminoglycosides

^{*2} BEL beta-lactamase | cephalosporins, penams, monobactams

*3 OXA beta-lactamase | carbapenems, cephalosporins, penams

^{*4} GES beta-lactamase | carbapenems, cephalosporins, penams

*5 Major facilitator superfamily (MFS) antibiotic efflux pump | phenicol

*6 Trimethoprim resistant dihydrofolate reductase dfr | diaminopyrimidines

*7 Macrolide esterase | Macrolides

^{*8} Small multidrug resistance (SMR) antibiotic efflux pump | disinfecting agents and antiseptics

2 Supplementary Figures

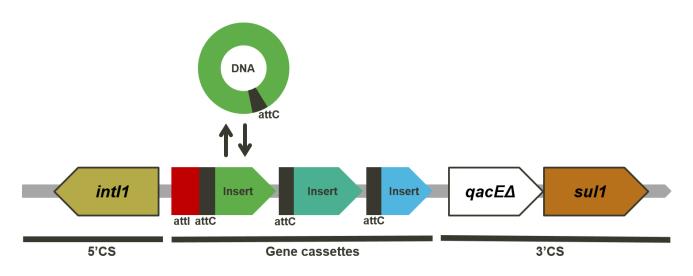


Figure S1. Structure of the Class 1 integron.

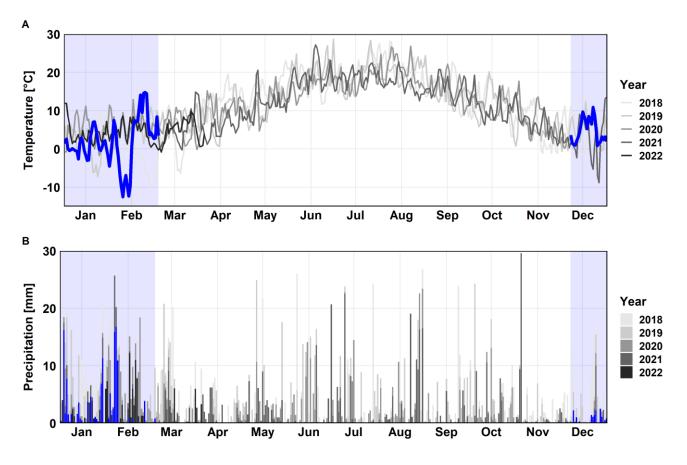


Figure S2. Temperature (A) and precipitation (B) in Wernigerode city from January 2018 until March 2022. Blue lines and bars are highlighting the data from December 2020 until March 2021. Data was received from the German weather monitoring station 5490 (https://opendata.dwd.de/).

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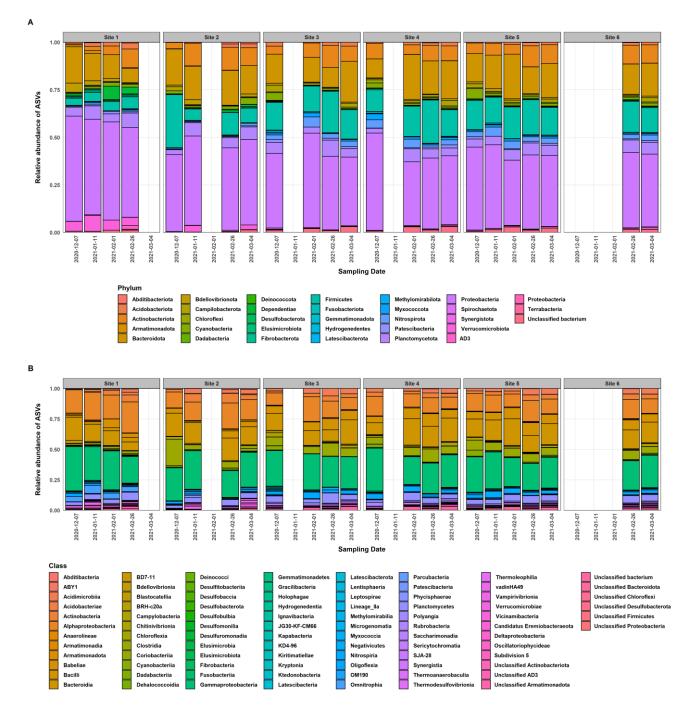


Figure S3. Microbial community structure at phylum (A) and class level (B) for each site, sorted by sampling date. For Sites 1 to 4, one outlier was removed in each data set before the analysis.

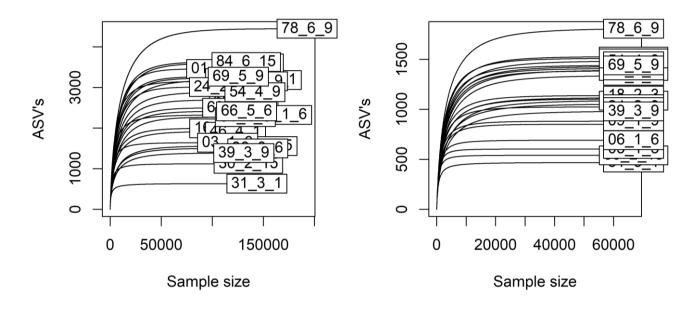


Figure S4. Rarefaction curves for 16S rRNA gene sequencing before (left) and after (right) removal of low abundance ASV's.

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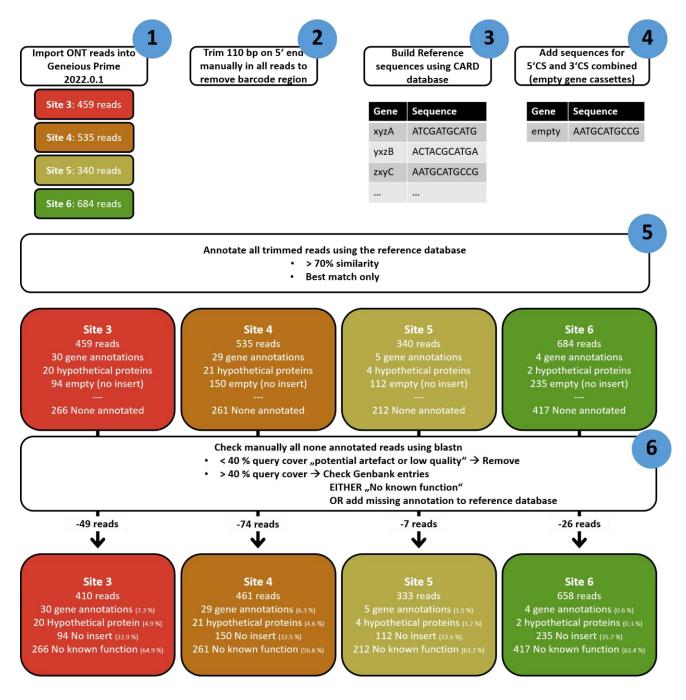


Figure S5. Workflow for the analysis of class 1 integron gene cassette inserts after ONT sequencing.

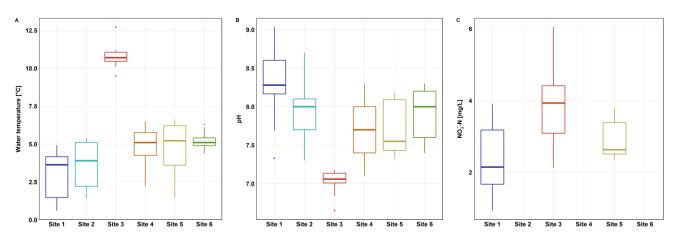


Figure S6. Physicochemical parameters measured at all Sites. Measurements were obtained from MOBICOS monitoring stations (site 1 and 5, https://www.ufz.de/index.php?en=39611), WWTP Silstedt (site 3, https://www.wahb.eu/) and manually using a SenTix41 probe (site 2, 4 and 6). Water temperature (A). pH (B) and NO3⁻-N (C). Distinct colours were used to differentiate between individual Sites.

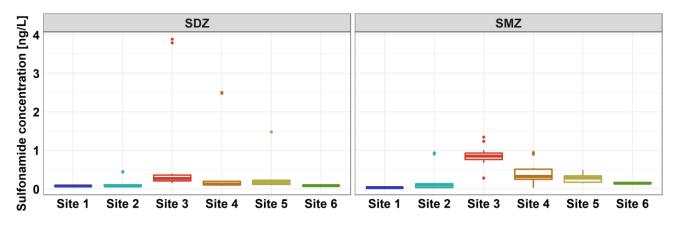


Figure S7. SDZ and SMZ concentrations for all six Sites measured by HPLC-MS/MS. The results of seven replicates are shown using box-whiskers-plots with the median represented by a horizontal line. Distinct colours were used to differentiate between individual Sites.

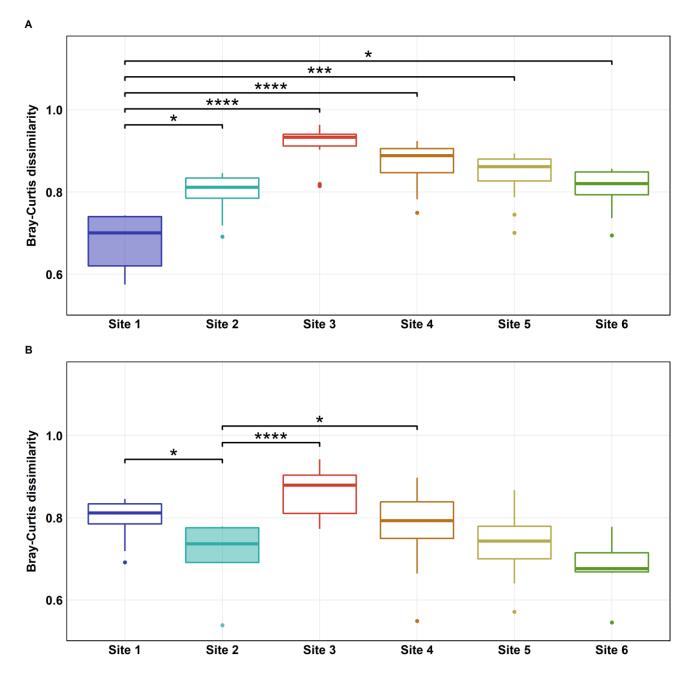


Figure S8. Bray Curtis dissimilarity in comparison to site 1 - pristine (A). Bray Curtis dissimilarity in comparison to site 2 - city (B). Significant differences between Sites are represented by asterisks (Dunn's test, * ≤ 0.05 , *** ≤ 0.001 , **** ≤ 0.0001). Distinct colours were used to differentiate between individual Sites.

3 References

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